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NUT/RB MICROSLIDE® TECHNICAL DOCUMENT





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NUT/RB

CODE: M-NUT/RB

USE

Isolation and differentiation of Gram (-) enteric bacilli. (**NUT**) Selective enumeration and cultivation of yeasts, molds, and Actinomycetes from food and other surfaces (**RB**).

APPLICATION

In total coliform testing (TCC), the coliform organisms tested for include: total coliform, fecal coliform, and E. coli (Escherichia coli). Detection of fecal coliforms (a subset of total coliforms) or Escherichia coli (a subset of fecal coliforms) can indicate the potential presence of waterborne pathogens associated with fecal contamination¹. Rose Bengal Agar is recommended in *Standard Methods* for the enumeration of yeasts and molds from food and water.

PADDLE AGARS



Side 1: Nutrient-TTC Agar (NUT) – (Color: Yellow) General purpose (relatively non-selective) medium, which will support the growth of a wide variety of organisms. Suitable for cultivation of both aerobes and anaerobes. Aerobic coliform bacteria can be detected by their ability to reduce the TTC dye to a red-colored formozan dye. Bacterial colonies appear as red dots on an otherwise yellow medium.

Note: Paddle color is normally LIGHT YELLOW when the NUT agar is cast (about pH 6.0). Some microorganism growth (even before colonies are OBSERVABLE) will shift the pH from an acidic to a more alkaline level (pH 7.0 or higher) – turning the agar a light green.

Side 2: Rose Bengal Agar (RB) – (Color: Pink) Selective medium for the enumeration of yeasts and molds.

*Note: Side 1 of each paddle is marked with an indented laser line.

STORAGE / EXPIRATION

Microslides[®] should be stored tightly sealed (unopened) in a cool, dry location at room temperature (18 - 25°C; 65 - 77°F). Temperature fluctuations may result in condensation settling at the bottom of the vial, although this does not affect culture properties, it could reduce the shelf-life or cause the agar to separate from the plastic paddle support. Refer to 'Best Before End date' (SEE: BBE stamped on vial).

Avoid sudden temperature changes. Shield from direct sunlight. Do not allow paddles to freeze. Do not store in a refrigerator (~44°F / 10°C) or at temperatures exceeding 80°F; 27°C. Refrigeration may result in water condensation. Discard if paddle agar appears oxidized (darkened from expected color) or if contaminants appear. Expiry applies to medium in its intact container when stored as directed.

¹ United States Pharmacopeial Convention. 2007. The United States pharmacopeia, 31st ed., Amended Chapters 61, 62, 111. The United States Pharmacopeial Convention, Rockville, MD.

AGAR VERIFICATION

These agars have been verified by <u>EMSL Analytical</u>, <u>Inc.</u> using *E. coli* and *E. faecalis* (NUT) and *P. commune* and *C. albicans* (RB) cultures. Documentation available upon request.

SAMPLING

SURFACE Sampling Protocol

- 1. Remove the paddle from the vial. Do not touch the agar surfaces.
- 2. To assure an accurate area recovery, contact the paddle to 20²cm of the surface by contacting the surface twice in separate 10²cm areas.
- 3. Replace paddle in vial.
- 4. Incubate.

LIQUID Sampling Protocol

DIRECT IMMERSION PROTOCOL - low viscous liquids

- 1. Mix liquid test sample.
- 2. Remove the paddle from the vial. Do not touch the agar surfaces.
- 3. When taking the sample:
 - a. Pour 40mL of the sample into the vial (to the printed horizontal fill line; see right). Dip the paddle into the 40mL volume liquid in the vial. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.



- b. Or dip the paddle into the sample directly. Maintain a contact time of at least 15 seconds (30 seconds optimal). Both agar surfaces must be completely contacted.
- 4. Allow excess fluid to drain off both paddle agar surfaces.
- 5. Replace paddle in vial.
- 6. Incubate.

SPREAD Protocol – high viscous liquids

- 1. Mix liquid test sample.
- 2. Remove paddle from vial. Do not touch the agar surfaces.
- 3. Holding the contact agar surface on a horizontal plane, deposit volume as a single drop approximately 1cm from the handle boundary (Figure 1).
- Position a sterile glass rod on the "handle" side of the drop and bring it into contact with the drop creating a meniscus. Drag the glass tube over the paddle agar surface.
- 5. Replace paddle in vial.
- 6. Incubate.



Figure 1

INCUBATION

Incubation of Paddle Growth	Incubation Temperature	Examine at:
Yeast / Mold	25 to 30°C	48 hours up to 120 hours (5 days)
Yeast / Mold	Room Temperature	Up to 7 days
Total Coliform / Bacteria	35 ± 2°C	24 to 48 hours
Total Coliform / Bacteria	Room Temperature	Up to 5 days

Note: Incubation of bacteria after 48 hours may produce confluent growth making enumeration more difficult.

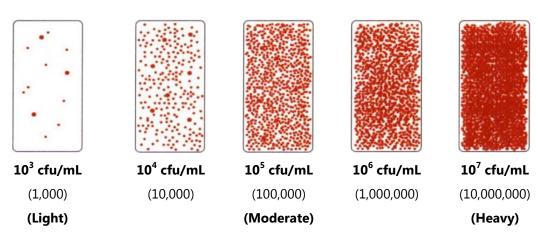
COLONY MEASURING

Each Microslide® paddle has molded media attachment points that are 4mm in length (point-to-point). This feature provides a useful guidepost to estimating nearby colony size.



ENUMERATION

Bacteria CFU/mL



Note: Estimation of lower counts is possible, but statistically difficult to justify. Use Light, Moderate and Heavy for Mold growth and surface testing.

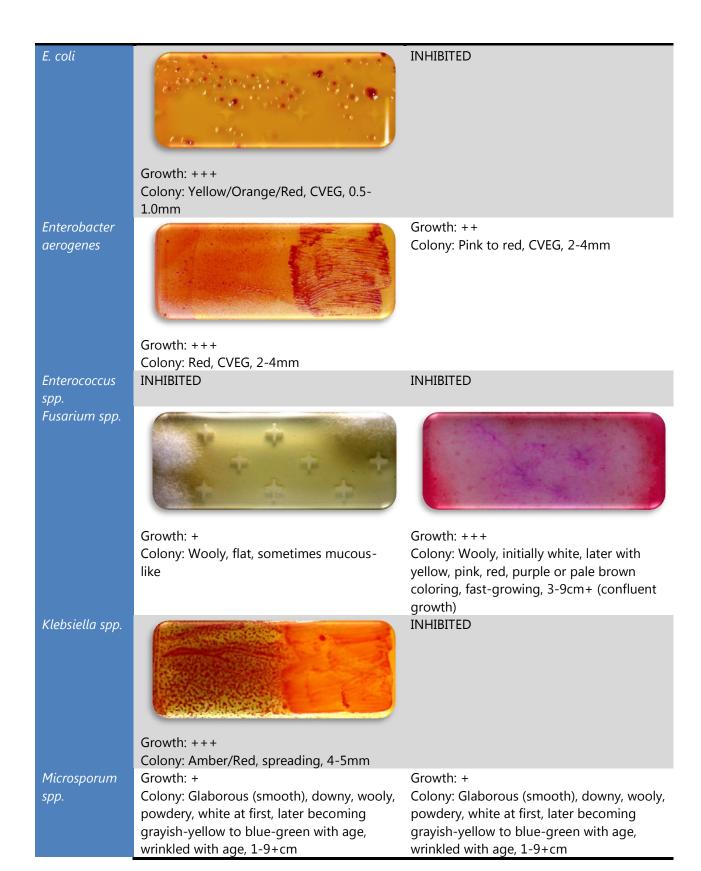
DISPOSAL

Make a 1:9 dilution of household bleach (5.25% sodium hypochlorite solution). Twist and remove Microslide[®] paddle from vial. Fill vial with 40mL diluted hypochlorite solution (to fill-line). Allow 15-minute contact time. Discard bleach solution. Replace paddle in vial and dispose. Alternatively, loosen cap and microwave for 30 seconds, autoclave, or incinerate.

IDENTIFICATION

Organism	Nutrient-TTC (NUT)	Rose Bengal (RB)
Actinomyces bovis Alternaria spp.	Growth: + Colony: Opaque/tan-grey, CVEG, 1-3mm Growth: + Colony: Downy to wooly; flat, grayish, short, aerial hyphae, later becomes greenish black or olive-brown with a light border, 3-9cm	Growth: ++ Colony: Opaque/tan-grey, CVEG, 1-3mm Growth: ++ Colony: Suede-like to woolly, initially white to yellow-orange, becoming black to olive- green or grayish, or grayish-green, umbonate with lighter center area,
	bolder, 3-9cm	condication (rings), fast-growing, 3-9cm+ (confluent growth)
Aspergillus niger		
	Growth: +++	Growth: +++
	Colony: Granular, jet black conidia with yellow/gray hyphae, 3-5++cm	Colony: Woolly and/or felt-like, forms a carpet, initially white later with jet black fruiting bodies (sporangia), fast-growing (4.5cm in 4 days), 3-9cm+ (confluent growth)
Aspergillus	Growth: +	Growth: +++
flavus	Colony: Granular to wooly, yellow, yellow- green, or yellow-brown, 3-9cm	Colony: Granular to wooly, yellow, yellow- green, or yellow-brown, 3-9cm+ (confluent growth)
Aspergillus	Growth: +	Growth: +++
fumigatus	Colony: Granular to cottony, blue-green, green-grey, or green-brown, 3-9cm	Colony: Felt-like, forms a carpet, initialy white to green or blue-green fruiting bodies, 3-9cm+ (confluent growth)
Aspergillus	Growth: +	Growth: +++
terreus	Colony: Granular, radially rugose (wrinkled), cinnamon buff/brown, 3-9cm	Colony: Granular, radially rugose (wrinkled), cinnamon buff/brown, 3-9cm+ (confluent growth)





Muccor spp.	Growth: + Colony: Wooly, fluffy (like cotton candy),	Growth: + Colony: Woolly, initially white, then white-
2	white at first, later becoming gray/yellow to blue-green with age, 2-5++cm	yellow to various shades of gray to green with lollipop fruiting bodies (sporangia), fast-growing, 3-9cm+ (confluent growth)
Penicillium chrysogenum (notatum)		
	Growth: ++ Colony: Granular, velvety/powdery, flat, initially white, then various shades of green-blue, green, or yellow-green, 3-5cm	Growth: ++ Colony: Granular, velvety/powdery, flat, initially white, then various shades of green-blue, green, or yellow-green, 3- 9cm+ (confluent growth)
Penicillium roqueforti		
	Growth: +	Growth: ++
	Colony: Granular, dull, green in coloar, arachnoid (with many spider web-like fibers) colony margins, 0.5-1.0cm	Colony: Granular, velvet-like, flat, initially white then various shades of green, bluegreen pigment, 3-9cm+ (confluent growth)
Penicillium	Growth: +	Growth: +++
digittum	Colony: Wooly, fluffy (like cotton candy), white at first, later becoming green with age, 3-9cm	Colony: Suede-like, woolly, initially white, then various shades of olive green, 3-9cm+ (confluent growth)
Pithomyces	Growth: +	Growth: +++
spp.	Colony: Powdery, pale/dark grey or brown pigment, 2-9++cm	Colony: Powdery, pale/dark grey or brown to olive green pigment, lighter outer ring with center bullseye, 2-9cm+ (confluent growth)
Proteus spp.	INHIBITED	INHIBITED

Pseudomonas aeruginosa



INHIBITED

Growth: +++

Colony: Red, irregular, spreading to

confluent, 2-4mm

Pseudomonas fluorescens



Growth: +++

Colony: Clear/colorless with grey/dark

center, translucent edges,

irregular/spreading to confluent, 2-4mm

Growth: +

Colony: Translucent, pinkish, or amber, irregular, raised, undulate, 2-4mm+

Rhizopus spp.



Growth: +++ Gro

Colony: Cottony, white to black/grey (black fruiting bodies), 2-9++cm

Growth: +++

Colony: Dense, cottony growth, initially white, turning to gray with black fruiting bodies (sporangia), fast-growing, 3-9cm+(confluent growth)

Saccharomyce cerevisiae

Salmonella



Growth: +++

Colony: Translucent to white or cream,

CVEG (may be dull), 0.1-0.5mm

(punctiform)
INHIBITED

Growth: ++

Colony: Creamy white to tan, spreading, circular, entire, raised to convex, glistening

surface, 5-8mm

Growth: +++

typhimurium Colony: I

Colony: Purple/pink, FED, 0.5-1.0mm



	green or yellow-green patches (rings), 2- 9++cm	green or yellow-green patches (rings), 3- 9cm+ (confluent growth)
Trichophyton	Growth: +	Growth: ++
spp.	Colony: Wooly with indented boarders, white to brown/tan pigment, 2-9++cm	Colony: Wooly , initially white with brownish/tan pigmentation, outer darker ring, 3-9cm+
Gram (+) Bacteria	PARTIAL TO COMPLETE INHIBITION	

GLOSSARY

CVEG	Convex, Entire, Glossy
FED	Full, Entire, Dull
Gram	Gram reaction